

ANTIOXIDANT ACTIVITY AND FLAVONOIDS OF *Stachys ocymastrum*

Hichem Lakhal,<sup>1</sup> Tarek Boudiar,<sup>1</sup> Ahmed Kabouche,<sup>1\*</sup>  
Souheila Laggoune,<sup>1</sup> Zahia Kabouche,<sup>1</sup> and Gulacti Topcu<sup>2</sup>

UDC 547.972

*Stachys* genus (Lamiaceae) has shown various activities such as anti-inflammatory [1], antimicrobial [2], and antioxidant [3] activities. Because of the various biological interests in the secondary metabolites (flavonoids, diterpenes, phenylethanoid glycosides) of this genus, we have made a phytochemical study of the antioxidant activity of the species *Stachys ocymastrum* (L.) Briq. [4].

Aerial parts of *Stachys ocymastrum* (L.) Briq. were collected from Djebel El-Ouahch-Constantine (North Eastern Algeria) in June 2005 during the flowering stage. A voucher specimen has been deposited in the Herbarium of the Department of Chemistry, University Mentouri-Constantine, and authenticated by Prof. G. De Belair (University of Annaba, Algeria).

Air-dried and powdered aerial parts (890 g) of *Stachys ocymastrum* were macerated in a methanolic solution (70%) at room temperature. The extract was concentrated under low pressure, diluted, and filtered, then successively extracted with petroleum ether, dichloromethane, ethyl acetate, and *n*-butanol.

Compound **1** was isolated as a yellow solid that precipitated from the ethyl acetate extract; then, the butanolic and the ethyl acetate extracts were concentrated under reduced pressure. The two extracts were combined and column chromatographed on Polyamid SC6 with a gradient of toluene–MeOH with increasing polarity, affording compound **2** from fraction F-75. Successive separations using preparative TLC on silica gel eluted with CH<sub>2</sub>Cl<sub>2</sub>–MeOH (9:1) led to compound **3** from fraction F-57, while paper chromatography (Whatman No. 3MM) eluted with AcOH–H<sub>2</sub>O (30:70) led to compound **4** from fraction F-70.

Compounds **1** and **2** were identified by <sup>1</sup>H NMR, <sup>13</sup>C NMR, Dept-135, <sup>1</sup>H–<sup>1</sup>H COSY, HMBC, HMQC, and MS, while compounds **3** and **4** were identified by UV and <sup>1</sup>H NMR as well as by direct comparison with literature data [6–9].

**Compound 1.** C<sub>30</sub>H<sub>26</sub>O<sub>12</sub>, mp 270°C. <sup>1</sup>H NMR (300 MHz, DMSO-d<sub>6</sub>, δ, ppm, J/Hz): 12.97 (1H, s, 5-OH), 7.94 (2H, d, J = 8.7, H-2', H-6'), 6.92 (2H, d, J = 8.7, H-3', H-5'), 6.83 (1H, s, H-3), 6.82 (1H, d, J = 1.8, H-8), 6.47 (1H, d, J = 1.8, H-6), 5.17 (1H, d, J = 6.9, H-1''), 4.47 (1H, d, J = 11.6, H-6''<sub>b</sub>), 4.16 (1H, dd, J = 11.9, 7.2, H-6''<sub>a</sub>), 3.84 (1H, m, H-5''), 3.35 (1H, m, H-2''), 3.33 (1H, m, H-3''), 3.26 (1H, m, H-4''), 7.49 (1H, d, J = 15.9, H-β), 6.67 (2H, d, J = 8.5, H-3''', H-5'''), 7.36 (2H, d, J = 8.5, H-2''', H-6'''), 6.32 (1H, d, J = 15.9, H-α). <sup>13</sup>C NMR (125 MHz, DMSO-d<sub>6</sub>, δ): 94.60 (C-8), 99.34 (C-6), 102.90 (C-3), 105.26 (C-10), 115.90 (C-3', C-5'), 120.86 (C-1'), 128.45 (C-2', C-6'), 156.80 (C-9), 161.02 (C-5), 161.28 (C-4'), 162.58 (C-7), 164.17 (C-2), 181.88 (C-4), 63.33 (C-6''), 69.87 (C-4'), 72.84 (C-2''), 73.70 (C-5''), 76.11 (C-3''), 99.37 (C-1''), 113.61 (C-α), 115.57 (C-3''', C-5'''), 124.78 (C-1'''), 130.00 (C-2''', C-6'''), 144.85 (C-β), 159.68 (C-4'''), 166.37 (C=O). Mass spectrum, *m/z*: 601.1 [M + Na]<sup>+</sup>, 623.1 [M – H + 2Na]<sup>+</sup>, 639.1 [M – H + Na + K]<sup>+</sup>. Characterized as apigenin 7-*O*-β-D-(6''-*O*-*p*-coumaroylglucopyranoside) [5].

**Compound 2.** C<sub>27</sub>H<sub>30</sub>O<sub>16</sub>. <sup>1</sup>H NMR (300 MHz, DMSO-d<sub>6</sub>, δ, ppm, J/Hz): 12.04 (1H, s, 5-OH), 8.35 (1H, s, 8-OH), 7.99 (2H, d, J = 8.8, H-2', H-6'), 6.96 (2H, d, J = 8.8, H-3', H-5'), 6.85 (1H, s, H-3), 6.65 (1H, s, H-6), 5.11 (1H, d, J = 7.5, H-1''), 3.74 (1H, m, H-6''<sub>b</sub>), 3.65 (1H, m, H-2''), 3.51 (1H, m, H-5''), 3.49 (1H, m, H-6''<sub>a</sub>), 3.49 (1H, m, H-3''), 3.36 (1H, m, H-4''), 4.91 (1H, d, J = 8, H-1'''), 3.87 (1H, m, H-3'''), 3.64 (1H, H-5'''), 3.52 (1H, H-6''<sub>b</sub>), 3.42 (1H, H-6''<sub>a</sub>), 3.34 (1H, m, H-4'''), 3.21 (1H, m, H-2'''). <sup>13</sup>C NMR (125 MHz, DMSO-d<sub>6</sub>, δ): 182.3 (C-4), 163.98 (C-2), 161.24 (C-4'), 152.35 (C-5), 151.13 (C-7), 144.24 (C-9), 128.56 (C-2', C-6'), 127.02 (C-8), 121.17 (C-1'), 115.91 (C-3', C-5'), 105.11 (C-10), 102.5 (C-3), 98.65 (C-6), 99.5 (C-1''), 81.17 (C-2''), 76.94 (C-3''), 75.59 (C-5''), 69.17 (C-4''), 60.4 (C-6''), 101.6 (C-1'''), 74.5 (C-5'''), 71.5 (C-2'''), 70.9 (C-3'''), 67.1 (C-4'''), 60.9 (C-6'''). Mass spectrum, *m/z*: 633.1 [M + Na]<sup>+</sup>, 655.1 [M – H + 2Na]<sup>+</sup>. Characterized as isoscutellarein 7-*O*-β-D-allopyranosyl-(1→2)-glucopyranoside [6, 7].

1) Laboratoire d'Obtention de Substances Therapeutiques (L.O.S.T), Faculte des Sciences Exactes, Universite Mentouri-Constantine, Campus Chaabat Ersas, 25000 Constantine, Algeria, e-mail: ahkabouche@yahoo.fr; 2) Department of Chemistry, Faculty of Science and Letters, Istanbul Technical University, 34469 Maslak, Istanbul, Turkey. Published in *Khimiya Prirodnykh Soedinenii*, No. 6, pp. 821–822, November–December, 2010. Original article submitted July 3, 2009.

**Compound 3.** C<sub>15</sub>H<sub>10</sub>O<sub>5</sub>, mp 327°C, characterized as luteolin [8].

**Compound 4.** C<sub>15</sub>H<sub>10</sub>O<sub>6</sub>, mp 347°C, characterized as apigenin [9].

Compound **1** was isolated for the first time from the genus.

**Antioxidant Activity.** The radical scavenging activity of the butanolic extract of *Stachys ocymastrum* (L.) Briq. was measured by the slightly modified method of Hatano [8]. The *n*-butanolic extract of *Stachys ocymastrum* (L.) Briq. exhibited good activity: IC<sub>50</sub> 6.77 ± 0.2 µg/mL compared with the reference (rutin: IC<sub>50</sub> 3.01 ± 0.2 µg/mL).

## ACKNOWLEDGMENT

The authors thank the National Health Research Agency, Oran, Algeria (ANDRS) and (DG/RSDT, MESRS), Algeria for financial support.

## REFERENCES

1. N. Maleki, A. Garjani, H. Nazemiyah, N. Nilfouroushan, S. Eftekhar, Z. Allameh, and N. Hasannia, *J. Ethnopharmacol.*, **75**, 213 (2001).
2. H. D. Skaltsa, C. Demetzos, D. Lazari, and M. Sokovic, *Phytochemistry*, **64**, 743 (2003).
3. A. Matkowski and M. Piotrowska, *Fitoterapia*, **77**, 346 (2006).
4. P. Quezel and S. Santa, *Nouvelle Flore de l'Algerie et des Regions Desertiques et Meridionales*, **1–2**, CNRS, Paris, 1963.
5. P. Venturella, A. Bellino, and M. L. Marino, *Phytochemistry*, **38**, 527 (1995).
6. A. Lenherr and T. J. Mabry, *Phytochemistry*, **26**, 1185 (1987).
7. M. A. El-Ansari, D. Barron, M. F. Abdalla, N. A. M. Saleh, and J. L. Le Quere, *Phytochemistry*, **30**, 1169 (1991).
8. T. J. Mabry, K. R. Markham, and M. B. Thomas, *The Systematic Identification of Flavonoids*, Springer, Berlin, 1970.
9. T. Hatano, H. Kagawa, T. Yasuhara, and T. Okuda, *Chem. Pharm. Bull.*, **36**, 2090 (1988).